## AANP Diagnostic Slide Session 2019 – Case #6 Julieann C. Lee and Joanna J. Phillips University of California San Francisco



Atlanta, Georgia June 8, 2019 The authors have nothing to disclose. The decedent was a 23-year-old woman.

There were early developmental delays.

Progressive motor regression began at age 10.

By her late teens she was almost entirely wheelchair bound, and fed primarily by a gastric tube.

She became increasingly nonverbal, with dystonia and ballistic movements.



Bilateral T1 hyperintense signal abnormalities within the caudate, putamen, globus pallidus, thalamus, hippocampus, red nucleus, and cortical spinal tract

#### Fresh brain weight: 745 grams

Coronal section with decreased bulk and coloration of globus pallidus Severely reduced volume of the corpus callosum and cerebral white matter





## Cortex 200x

## NeuN 200x



## White matter, 100x



# Globus pallidus, 200x



Pineal gland

Dentate gyrus

DRG

## **Differential diagnosis?**

Additional immunohistochemical or molecular evaluation?



Prussian blue iron stain, basal ganglia

Special staining for iron demonstrates abnormal deposition in rare cells, and highlights a subset of the eosinophilic deposits



Electron microscopy of the eosinophilic inclusions and globules showed electron dense granular material, some of which was membrane bound with peripherally displaced chromatin, while other large collections were apparently not directly associated with cellular structures. The inclusions and globules contained collections of material compatible with ferritin.

# The eosinophilic globules and inclusions were immunopositive for Ferritin



### **FINAL AUTOPSY DIAGNOSIS**

#### Neurodegeneration with features characteristic of a neuroferritinopathy

A. Eosinophilic intranuclear and intracytoplasmic inclusion bodies, apparent extracellular deposits, and patchy iron deposition.

B. Diffuse and extensive involvement of brain and spinal cord with severe involvement and patchy destruction of basal ganglia, red nucleus, substantia nigra, and white matter of cerebral cortex and cerebellum.

C. Severe atrophy/hypoplasia of the cerebellar vermis and brain stem.

D. Hydrocephalus ex vacuo.

D. Eosinophilic intranuclear inclusion bodies in multiple organs including skeletal muscle, Schwann cells in peripheral nerve, endothelial cells, adventitia, the mesenchyma of the heart, the adrenal cortex, occasional hepatocytes in the liver, and the collecting ducts and tubules of the kidney.



Arosio et al. Biochem J. 2015 Nov 15;472(1):1-15.



Ferritins: heteropolymers composed of 24 light (FTL) and heavy (FTH1) polypeptide subunits in variable proportions

Ferritins are involved in iron binding, metabolism, detoxification, and storage

#### Ferritin light chain FTL mutations

cause autosomal dominant neurodegenerative disease with cellular inclusions and ferritin accumulations "Hereditary Neuroferritinopathy"

Curtis et al. Nat Genet. 2001 Aug;28(4):350-4.



Purkinje cell with cytoplasmicExtracellular putamenGFAP + astrocyte withferritin + inclusionsdepositsvacuolated nucleus





Inclusions can be seen within neurons and glia

Both cytoplasmic and nuclear inclusions occur

Extracellular deposits are also found

Manusco *et al.* J Neuropathol Exp Neurol. 2005

Vidal *et al.,* J Neuropathol Exp Neurol 2004



Iron- and ferritin-positive deposits were present throughout the forebrain and cerebellum,

these being mainly extracellular

but also showing colocalization with microglia and oligodendrocytes as well as with neurons, notably in the globus pallidus.

Curtis *et al.,* Mutation in the gene encoding ferritin light polypeptide causes dominant adult-onset basal ganglia disease. Nat Genet. 2001 Aug;28(4):350-4. PMID: 11438811.



Eosinophilic nuclear inclusions are also present in other organs "Hereditary ferritinopathy" is not exclusively a neuroferritinopathy # 606159

# NEURODEGENERATION WITH BRAIN IRON ACCUMULATION 3; NBIA3

Alternative titles; symbols

#### NEUROFERRITINOPATHY BASAL GANGLIA DISEASE, ADULT-ONSET

**Phenotypic Series** 



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#### **Phenotype-Gene Relationships**

		Dhanatura		Dhonotuno		Gene/Locus
Location	Phenotype	MIM number	Inheritance	mapping key	Gene/Locus	MIM number
19q13.33	Neurodegeneration with brain iron accumulation 3	606159	<u>AD</u>	<u>3</u>	FTL	134790

PheneGene Graphics •

**TEXT** 

A number sign (#) is used with this entry because of evidence that this form of neurodegeneration with brain iron accumulation (NBIA), here designated 'NBIA3,' is caused by heterozygous mutation in the FTL gene (134790) on chromosome 19q13. See NOMENCLATURE section.

For a general phenotypic description and a discussion of genetic heterogeneity of NBIA, see NBIA1 (234200).

#### Description

Neurodegeneration with brain iron accumulation is a genetically heterogeneous disorder characterized by progressive iron accumulation in the basal ganglia and other regions of the brain, resulting in extrapyramidal movements, such as parkinsonism and dystonia. Age at onset, cognitive involvement, and mode of inheritance is variable (review by Gregory et al., 2009).

Neurodegeneration with brain iron accumulation (NBIA)

> NBIA-type 1 PANK2 mutation

NBIA-type 2 PLA2G6 mutation

> NBIA-type 3 FTL mutation

Details on NBIA types 1-8 can be found at OMIM (Online Mendelian Inheritance in Man)



## Hereditary Neuroferritinopathy

- Ferritin light chain *FTL* mutations cause AD neurodegenerative disease

- Inclusions within neurons and glia, and extracellular deposits

- Nuclear inclusions are also found in cells outside the CNS (ex. hepatocytes and renal tubule epithelium)
- A type of Neurodegeneration with Brain Iron Accumulation (NBIA)





## References

1. Vidal R, Ghetti B, Takao M, Brefel-Courbon C, Uro-Coste E, Glazier BS, Siani V, Benson MD, Calvas P, Miravalle L, Rascol O, Delisle MB. Intracellular ferritin accumulation in neural and extraneural tissue characterizes a neurodegenerative disease associated with a mutation in the ferritin light polypeptide gene. J Neuropathol Exp Neurol. 2004 Apr;63(4):363-80. PMID: 15099026.

2. Mancuso M, Davidzon G, Kurlan RM, Tawil R, Bonilla E, Di Mauro S, Powers JM. Hereditary ferritinopathy: a novel mutation, its cellular pathology, and pathogenetic insights. J Neuropathol Exp Neurol. 2005 Apr;64(4):280-94. PubMed PMID: 15835264.

3. Curtis AR, Fey C, Morris CM, Bindoff LA, Ince PG, Chinnery PF, Coulthard A, Jackson MJ, Jackson AP, McHale DP, Hay D, Barker WA, Markham AF, Bates D, Curtis A, Burn J. Mutation in the gene encoding ferritin light polypeptide causes dominant adult-onset basal ganglia disease. Nat Genet. 2001 Aug;28(4):350-4. PMID: 11438811.

4. Vidal R, Miravalle L, Gao X, Barbeito AG, Baraibar MA, Hekmatyar SK, Widel M, Bansal N, Delisle MB, Ghetti B. Expression of a mutant form of the ferritin light chain gene induces neurodegeneration and iron overload in transgenic mice. J Neurosci. 2008 Jan 2;28(1):60-7. PMID: 18171923.

5. Chinnery PF, Crompton DE, Birchall D, Jackson MJ, Coulthard A, Lombès A, Quinn N, Wills A, Fletcher N, Mottershead JP, Cooper P, Kellett M, Bates D, Burn J. Clinical features and natural history of neuroferritinopathy caused by the FTL1 460InsA mutation. Brain. 2007 Jan;130(Pt 1):110-9. PMID: 17142829.

6. Arosio P, Carmona F, Gozzelino R, Maccarinelli F, Poli M. The importance of eukaryotic ferritins in iron handling and cytoprotection. Biochem J. 2015 Nov 15;472(1):1-15. PMID: 26518749.

7. Garringer HJ, Irimia JM, Li W, Goodwin CB, Richine B, Acton A, Chan RJ, Peacock M, Muhoberac BB, Ghetti B, Vidal R. Effect of Systemic Iron Overload and a Chelation Therapy in a Mouse Model of the Neurodegenerative Disease Hereditary Ferritinopathy. PLoS One. 2016 Aug 30;11(8):e0161341. PMID: 27574973.